carry out appraisals and guide the career development and training programme of research staff.

**Conclusion** The Global Health Network has created a flexible method and set of tools (PDS) to support researchers and teams to document their professional career and core competencies. It enables individuals and groups to easily identify and track capacity development; an essential requirement for conducting effective health research.

**Background** Improvement of the diagnosis of smear-negative pulmonary tuberculosis (PTB) patients and identification of possible immune factors associated with the negative result of sputum will enable early and accurate diagnosis of smear-negative PTB. This study aimed to measure the Mycobacterium load in sputum samples of smear-negative patients and identify cytokines markers associated with smear-negative active pulmonary tuberculosis.

**Methods** Sputum and heparinised blood samples were collected from 40 smear-negative, 40 smear-positive PTB patients and 21 healthy controls. All sputum samples were analysed by direct ZN stain and conventional PCR to confirm the infection and characterise the bacteria. The load of bacteria in sputum samples was measured using real-time PCR. Blood samples were stimulated with sonicated MTB H37Rv. TH1 (TNF-α, IFN-γ, IL-1β) and TH2 (IL-10) cytokines were measured using ELISA technique.

**Results** Eight patients were grade 3+, 23 were grade 2+, 9 were grade 1+ and 40 were negative on smear. 87.5% of smear-negative patients were positive by PCR. Smear-negative PTB patients produced high concentration of IFN-γ compared with smear-positive. IL-10 and TNF-α concentrations were significantly lower in smear-negative compared with smear-positive. IL-1β was not significantly different between smear-negatives and smear-positives. Both smear-negative and smear-positive samples produced significantly high IL-10 and TNF-α cytokine compared with the healthy controls, while IFN-γ production was significantly lower in MTB patients. A highly significant correlation between MTB load and cytokines was detected. The mean concentration of IFN-γ was higher in stimulated blood samples of patients with lower bacterial load. In contrast, IL-10 and TNF-α concentration were higher in patients with high bacterial load. The TNF-α and IL-1β were good biomarkers for diagnosis of smear-negatives.

**Conclusion** Smear-negative PTB produced high TH1 cytokine and low regulatory cytokine compared to smear-positive.

**Background** Most research consortia and multicentre trials include capacity strengthening as one of their objectives. Activities are included in annual reports but the overall success or otherwise of these is hard to evaluate.

**Methods** The training and capacity building work package in the EDCTP2-funded ScreenTB Consortium includes support for the individual career development of young researchers. We have made mentoring the central activity, building on what we had learnt in the previous AE-TBC Consortium. We carried out formal training sessions and provided dedicated time-slots for meetings of mentee and mentor during annual meetings. We also introduced the concept of personal development plans through presentations and small group work.

**Results** Formal timetabled presentations and mentoring sessions have helped make capacity strengthening work. Challenges and solutions have been identified in group sessions and will be presented. This has enabled us to evaluate what works well and what is more challenging, when including capacity strengthening activities within a consortium with 5 African and 3 European partners.

**Conclusion** Dedicated time and commitment are required to make capacity strengthening work, but when it does, mentoring and personal development planning can provide both African and European researchers with an impartial opportunity to find solutions to their current challenges and to discuss their longer-term goals.